

# Lyo NZYSupreme qPCR Probe Master Mix (2x)

## Catalogue number:

MB41701, for 1,5 ml (150 x 20 µL)

## Description

Lyo NZYSupreme qPCR Probe Master Mix (2x) is an optimized and highly efficient freeze-dried reaction mixture developed for real-time PCR. This master mix was engineered with a dual hot-start enzyme control mechanism to provide the highest detection sensitivity. In addition, the latest developments in PCR enhancers have been incorporated in the Lyo NZYSupreme qPCR Probe Master Mix, including buffer chemistry and incorporation of highly robust engineered enzymes. This master mix does not contain ROX and it was specifically developed for probe-detection technology, including molecular beacons. For qPCR instruments that require this reference dye, we indicate a concentration range for your assay according to the table shown in the section "ROX reference dye" (ROX not provided). Lyo NZYSupreme qPCR Probe Master Mix (2x) is provided as a simple-to-use, stabilized 2x reaction mixture that includes all components for quantitative PCR, except sample DNA, primers, probe and water.

## Shipping Conditions

The product can be shipped in a range of temperatures from room temperature to dry ice.

## Storage Conditions

Although the product components provided are stable at room temperature, upon arrival they should be stored at -85 °C to -15 °C in a freezer without defrost cycles in order to guarantee maximal shelf life. Short-term storage for 1-2 weeks at room temperature will not compromise the performance of the product. Once the lyophilized master mix has been resuspended using the buffer provided, it should be stored at -85 °C to -15 °C. Minimize the number of freeze-thaw cycles by storing in working aliquots. The master mix will remain stable till the expiry date if stored as specified.

## Pack components

Component	MB41701
Lyo NZYSupreme qPCR Probe Master Mix (2x)	3 tubes with lyophilized master mix
qPCR master mix reconstitution buffer	1.75 mL

## Preparation of Lyo NZYSupreme qPCR Probe Master Mix

Reconstitute each tube of Lyo NZYSupreme qPCR Probe Master Mix with 525 µL of the qPCR master mix reconstitution buffer

provided. Complete hydration of the mixture might take 2-3 min. Mix gently by flicking the tube, and spin down. Do not replace the reconstitution buffer with water or any other buffer. The master mix is then ready to use as a 2x qPCR master mix.

## Compatible real-time PCR instruments

The master mix is compatible with real-time PCR instruments that do not require a passive reference signal for data normalization. It has been optimized to be compatible with the following real-time PCR instruments:

### Bio-Rad®:

CFX96™; CFX384™; iCycler®; iQ™5; Opticon™; Opticon™ 2

### Qiagen (Corbett):

Rotor-Gene™ 3000; Rotor-Gene™ 6000 & Rotor-Gene™ Q

### Roche:

Lightcycler® 96; Lightcycler® 480 & Lightcycler® Nano

### Applied Biosystems (with optional ROX addition):

7000; 7300; 7700; 7900; 7900HT; 7900HT FAST; StepOne™ & StepOne™plus; 7500; 7500 FAST; QuantStudio™ 6, 7, 12k Flex & ViiA7™

## Protocol

The following protocol serves as a general guideline and a starting point for any qPCR procedure. Optimal reaction conditions (incubation times and temperatures, concentration of template DNA) may vary and, in particular conditions, may require further optimization.

**qPCR reaction set-up:** the given volumes are based on a standard 20 µL final reaction mix which can be scale adjusted.

Lyo NZYSupreme qPCR Probe Master Mix (2x) <sup>(*)</sup>	10 µL	1x
10 µM forward primer	0.8 µL	400 nM <sup>(**)</sup>
10 µM reverse primer	0.8 µL	400 nM <sup>(**)</sup>
10 µM probe	0.2 µL	100 nM <sup>(**)</sup>
Template	up to 8 µL	-
Nuclease-free water	up to 20 µL	-

<sup>(\*)</sup> The master mix does not contain ROX, but, if required, addition of this internal passive reference dye can be conducted in a separate step. The final concentration will vary according to the qPCR instrument used. Please follow instructions described in the section "ROX reference dye" below.

<sup>(\*\*)</sup> See section of "General considerations" below for more details about primers and probes final concentrations in the reaction.

**Testing and Ct values:** When comparing Lyo NZYSupreme qPCR Probe Master Mix (2x) with a mix from another supplier we strongly recommend amplifying from a 10-fold template dilution series. Loss of detection at low template concentration is the only direct measurement of sensitivity. An early Ct value is not an indication of good sensitivity, but rather an indication of speed.

**ROX reference dye:** Lyo NZYSupreme qPCR Probe Master Mix (2x) is compatible with most thermocyclers available in the market and can include ROX passive reference dye (not provided) to normalize non-PCR-related fluctuations in fluorescence. If ROX addition is required for your qPCR platform, an optimal quantity of this dye should be included in your master mix. The recommended amount of ROX for the most common qPCR instruments is stated in the table below:

qPCR instrument	Volume of ROX per 525 µL of 2x master mix (2x) (*)
Applied Biosystems:7000/7300/7700/7900/7900HT/7900HT FAST/StepOne™/ StepOne™plus	10-15 µL (500-725 nM)
Applied Biosystems:7500/7500FAST/QuantStudio™ 5, 6, 7, 12k Flex/ViiA7™	1-2 µL (50-100 nM)
Bio-Rad®: CFX96™/CFX384™/iCycler®/iQ™5/Opticon™/ Opticon™ 2 Qiagen: Rotor-Gene™ 3000/6000/Q	Not required

(\*) For different volumes please scale-up or scale-down the volume of ROX accordingly. Please notice that ROX is not provided by NZYTech.

### Suggested thermal cycling conditions

Lyo NZYSupreme qPCR Probe Master Mix (2x) was optimized for the amplification of DNA fragments up to 200 bp under different real-time PCR cycling conditions. The table below displays a standard set-up optimized on a number of platforms. However, these conditions may be adapted to suit different machine-specific protocols.

Cycles	Temp.	Time	Notes
1	*95 °C	*2-5 min	Polymerase activation
40	95 °C	5 sec	Denaturation
	60 °C	**30 sec – 50 sec	Annealing/Extension

\* 2 min for cDNA, up to 3 or 5 min for genomic DNA.

\*\* Up to 50 s may be necessary for multiplexing with more than two probes.

### General considerations

In order to prevent any DNA contamination, we recommend that users have independent areas for reaction set-up, PCR amplification and any post-PCR gel analysis. It is essential that any tubes containing amplified PCR product are not opened in the PCR set-up area.

**Primers and probe:** these guidelines refer to the design and set-up of TaqMan probe-based PCR. Please refer to the relevant literature when using other probe types. The specific amplification, yield and overall efficiency of any real-time PCR can be critically affected by the sequence and concentration of the probes and primers, as well as by the amplicon length. We strongly recommend taking the following points into consideration when designing and running your real-time PCR experiment:

- Primers should have a melting temperature ( $T_m$ ) of approximately 60°C. The probe  $T_m$  should be approximately 10 °C higher than that of the primers
- The fragment should be between 80-200 bp length and not superior to 300 bp
- Final primer concentration of 400 nM is suitable for most probe-based reactions. However, to determine the optimal concentration we recommend titrating in the range 0.2-1 µM. The forward and reverse primers concentration should be equimolar

- A final probe concentration of 100 nM is suitable for most applications; we recommend that the final probe concentration is at least two-fold lower than the primer concentration; to determine the optimal concentration we recommend titrating in the range 0.1-0.25 µM;
- For multiplex qPCR, probe concentrations in excess of 100 nM can result in cross channel fluorescence.

**Template:** It is important that the DNA template is purified and concentrated according to conventional nucleic acid clean up procedures. In addition, templates must be devoid of any contaminating PCR inhibitors (e.g., EDTA). The recommended amount of template for PCR is dependent upon the type of DNA used. Please consider the following points when using genomic DNA or cDNA templates:

- **Genomic DNA:** use up to 1 µg of genomic DNA in a single PCR. We recommend using NZY Tissue gDNA Isolation kit (Cat. No. MB135) for high yield and purity from both prokaryotic and eukaryotic sources.
- **cDNA:** the optimal amount of cDNA to use in a single PCR depends upon the copy number of the target gene; we suggest using 100 ng cDNA per reaction. However, this amount may be adjusted to a more appropriate concentration. We suggest using NZY First-Strand cDNA Synthesis Kit (Cat. No. MB125) for reverse transcription of purified RNA. To obtain high yield of highly purified RNA we suggest using NZY Total RNA Isolation Kit (Cat. No. MB134).

**MgCl<sub>2</sub>:** It is not necessary to supplement the mix with MgCl<sub>2</sub> as Lyo NZYSupreme qPCR Probe Master Mix (2x) already contains an optimized concentration of MgCl<sub>2</sub>.

**PCR controls:** The reliability of the data may be affected by the presence of contaminating DNA. It is strongly recommended to include a no-template control reaction in the qPCR design, replacing template DNA/cDNA with nuclease-free PCR-grade water (DEPC-treated Water, Cat. No. MB43701). When performing a two-step RT-PCR, set up a no-RT control as well as a no-template control for the PCR.

### Quality control assays

#### Genomic DNA contamination

The product must comply with internal standards of DNA contamination as evaluated through real-time qPCR.

#### Nuclease assays

To test for DNase contamination, 0.2-0.3 µg of pNZY28 plasmid DNA are incubated with the master mix for 14-16 h at 37 °C. To test for RNase contamination, 1 µg of RNA is incubated with the master mix for 1 h at 37 °C. Following incubation, the nucleic acids are visualized on a GreenSafe-stained agarose gel. There must be no visible nicking or cutting of the nucleic acids.

#### Functional assay

Lyo NZYSupreme qPCR Probe Master Mix is extensively tested for activity, processivity, efficiency, sensitivity and heat activation.

### Certificate of Analysis

Test	Result
Genomic DNA contamination	Pass
Nuclease assays	Pass
Functional assay	Pass

Approved by:



Patrícia Ponte  
Senior Manager, Quality Systems

*For research use only.*



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